DOI: 10.1002/adsc.200800129

Chemoenzymatic Syntheses of Tumor-Associated Carbohydrate Antigen Globo-H and Stage-Specific Embryonic Antigen 4

Zhen Wang,[a] Michel Gilbert,[b] Hironobu Eguchi,[c] Hai Yu,[d] Jiansong Cheng,[d] Saddam Muthana,[d] Luyuan Zhou,[a] Peng George Wang,[c] Xi Chen,[d] and Xuefei Huang[a,*]

[a] Department of Chemistry, The University of Toledo, 2801 W. Bancroft Street, MS 602, Toledo, Ohio 43606, USA
Fax: (+1)-419-530-4033; e-mail: Xuefei.huang@utoledo.edu
[b] National Research Council Canada, Institute for Biological Sciences, Glycobiology Program, 100 Sussex Drive, Ottawa, ON K1A 0R6, Canada
[c] The Ohio State University, Departments of Biochemistry and Chemistry, 484 West 12th Avenue, Columbus, OH 43210, USA
[d] Department of Chemistry, University of California, One Shields Avenue, Davis, CA USA

Received: February 29, 2008; Published online: April 11, 2008

Dedicated to Prof. Chi-Huey Wong on the occasion of his 60th birthday for his ground breaking contributions to chemistry.

Supporting information for this article is available on the WWW under http://asc.wiley-vch.de/home/.

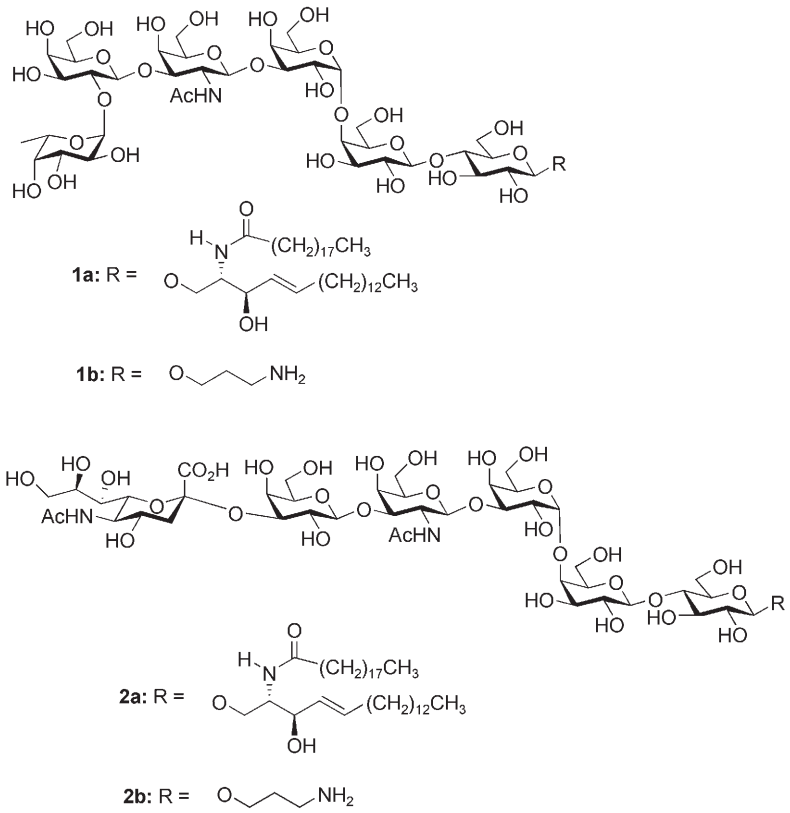
Abstract: Gangliosides have attracted much attention due to their important biological properties. Herein we report the first chemoenzymatic syntheses of two globo series of ganglioside oligosaccharides, Globo-H **1** and stage-specific embryonic antigen-4 (SSEA-4) **2**. The common precursor SSEA-3 pentasaccharide for these two compounds was assembled rapidly using the pre-activation-based one-pot glycosylation method. The stereoselectivity in forming the 1,2-*cis* linkage in SSEA-3 was attributed to a steric buttressing effect of the donor rather than electronic properties of the glycosyl donors. SSEA-3 was then successfully fucosylated by the fucosyltransferase WbsJ and sialylated by sialyltransferases CST-I and PmST1 producing Globo-H and SSEA-4, respectively.

Keywords: carbohydrates; chemoenzymatic synthesis; natural products; oligosaccharides; stereoselectivity

Introduction

The globo series of gangliosides, including Globo-H **1** and stage-specific embryonic antigen-4 (SSEA-4) **2**, possess the β-GalNAc-(1→3)-α-Gal-(1→4)-Gal oligosaccharide moiety.[1,2] This family of glycolipids has attracted much attention due to their roles as tumor-associated antigens[2] and as receptors for bacteria, viruses and toxins, which present excellent targets for novel therapeutics design.[3–5] For example, Globo-H has been found to be over-expressed on a variety of human cancer cells, including breast cancer, prostate cancer, ovarian cancer, and lung carcinomas.[6,7] Immunotherapy using Globo-H hexasaccharide as a cancer vaccine[8] has received encouraging preliminary results against breast cancer, which is now undergoing phase II clinical trials.[9–11] SSEA-4 is believed to be involved in bacterial and viral infections, serving as receptors for uropathogenic *Escherichia coli*[12–14] and human parvovirus B19.[15] In addition, SSEA-4 is a marker for human cancer cells[16–19] and the expression level of SSEA-4 shows clear correlation with metastasis potential and malignancy of renal cell carcinoma.[20–23]

Due to their biological importance and the difficulties in accessing these complex oligosaccharides from natural sources, great efforts have been devoted to chemical synthesis.[1,24] Globo-H hexasaccharide has been assembled by a variety of methods including the glycal strategy,[25] the trichloroacetimidate method,[26] two-directional glycosylation,[27] automated solid-phase synthesis,[28] the reactivity-based one-pot method[29] as well as the pre-activation-based one-pot method.[30] With the presence of the sialic acid unit at its non-reducing end, SSEA-4 presents additional synthetic challenges. Although much progress has been



made in chemical sialylation,[31,32] it still remains one of the most difficult glycosidic linkages to synthesize. The carboxylic acid moiety in sialic acid also requires additional consideration for protective group compatibility in synthetic design.[33] To date, SSEA-4 has only been assembled twice by the Hasagawa[34] and Schmidt/Garegg groups.[35] The successful completion of these highly challenging structures serves as highlights of the power of modern carbohydrate synthetic methodologies. However, despite these accomplishments, there is a continual need to further improve synthetic efficiencies of these molecules.

An attractive alternative to chemical synthesis is enzymatic synthesis.[36,37] With their high regio- and stereoselectivities, glycosyltransferases can facilitate the formation of glycosidic bonds under aqueous conditions without relying heavily on protective groups. However, enzymatic activities can be highly dependent upon structures of both the glycosyl donor and the acceptor. An effective enzymatic synthesis may require the preparation and screening of an array of enzymes, which can be time-consuming. Thus, chemoenzymatic synthesis, through the combination of the power of enzymatic synthesis with the flexibility of chemical synthesis, presents a potent approach to access complex oligosaccharides. Several gangliosides such as GM3,[38] GM2,[39] GM1,[39] GD1a,[39] Gb3[40] and SSEA-3[41] have been synthesized chemoenzymatically. Herein, we report our studies on chemoenzymatic syntheses of Globo-H **1b** and SSEA-4 **2b**, both of which contain aminopropyl side chains and can be conjugated to protein carriers for future immunological studies.

Results and Discussion

Retrosynthetically, we envision that both Globo-H and SSEA-4 can be accessed by enzymatic glycosylation of pentasaccharide SSEA-3 **3**, which is also a tumor-associated carbohydrate antigen.[42] The pentasaccharide **3**, in turn, can be chemically assembled from disaccharide **5**, galactoside **6**, and lactoside **7** (Figure 1). Based on our previous studies,[30] the presence of electron-withdrawing benzoyl moieties on the lactoside acceptor drastically reduced the glycosylation yield. Thus, the electron-donating benzyl groups are selected as protective groups masking hydroxy groups not undergoing glycosylation in order to enhance the nucleophilicity of the acceptors as well as to simplify deprotection procedures. A side effect of using benzyl groups is that building block galactoside **6** possesses high anomeric reactivity (armed)[43] as a glycosyl donor with its multiple electron-donating protective groups.[36] Therefore, in chemoselective glycosylation of thiogalactoside **6** by the thioglycoyl

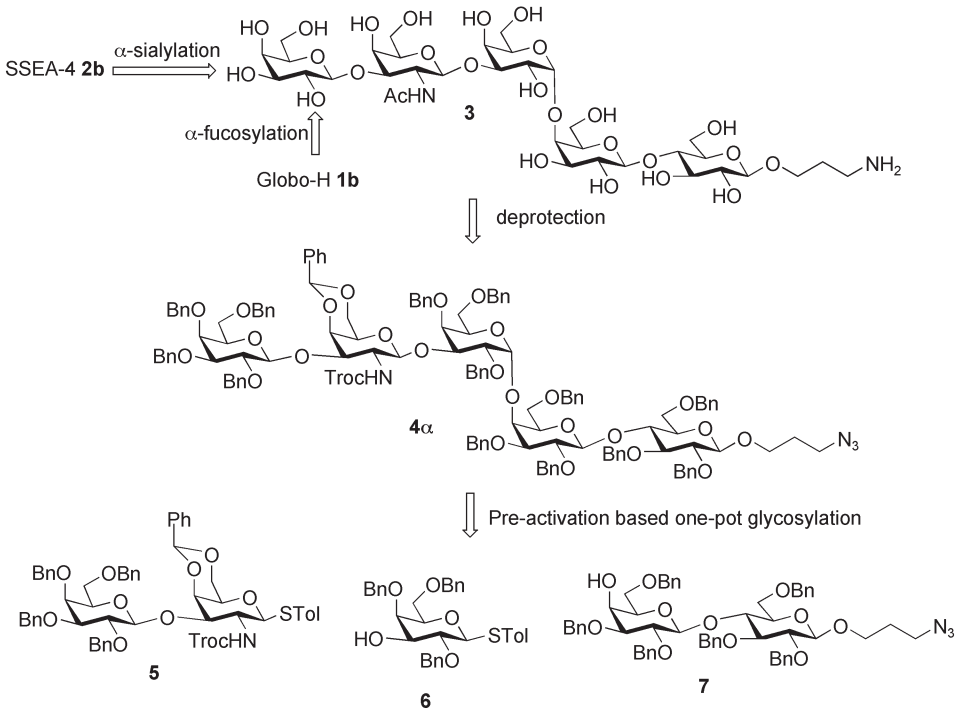


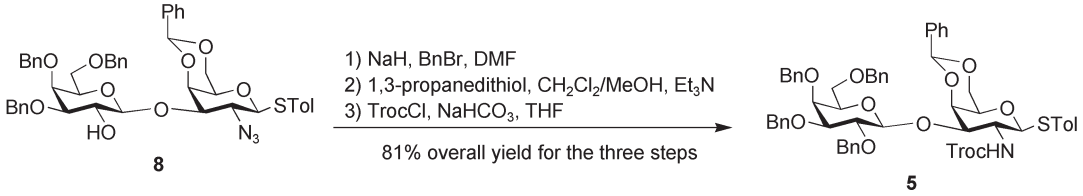
Figure 1. Retrosynthetic analysis.

donor **5**, caution must be taken to prevent the undesired premature activation of galactoside **6**. This can be achieved by pre-activating[44–48] disaccharide **5** first with a stoichiometric promoter, generating a reactive intermediate. Upon complete donor activation, addition of the bifunctional galactoside **6** building block to the reaction mixture will produce the desired trisaccharide. As donor activation and acceptor addition occur as two distinct steps, even though the acceptor **6** has high anomeric reactivity, it will not be activated by the promoter.

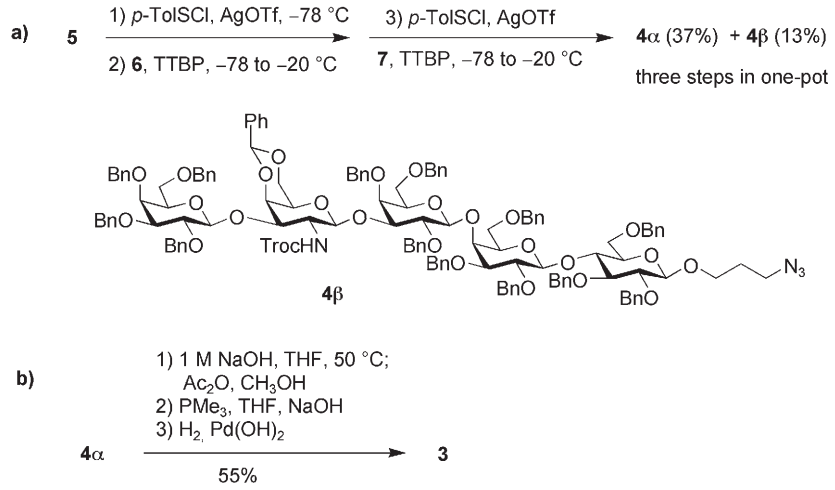
For the pre-activation-based glycosylation approach, we have extensively used the *p*-TolSCl/AgOTf promoter system,[30,44,49–52] which generates *p*-TolSOTf *in situ*. *p*-TolSOTf is a powerful thiophilic agent, capable of activating thioglycosides even with very low anomeric reactivities.[48,53,54] Moreover, the side product *p*-tolyl disulfide produced from pre-activation is not electrophilic, thus will not activate the thioglycoside acceptor. Alternatively, reagent combinations such as benzenesulfinylpiperidine/triflic anhydride (Tf_2O),[55] *S*-(4-methoxyphenyl) benzenethiosulfinate/Tf_2O,[56] benzenesulfinylmorpholine/Tf_2O,[57] and diphenyl sulfoxide/Tf_2O[58] have also been used as thioglycoside promoters for pre-activation reactions. However, electrophilic side products are typically generated with these promoters, which often require the addition of a quencher such as triethyl phosphite at the end of the reaction to prevent undesired acceptor activation.[45,59] This renders it difficult to carry out multiple sequential glycosylations in the same reaction flask without intermediate purification.

Disaccharide donor **5** was prepared from disaccharide **8**[30] starting with treatment of sodium hydride and benzyl bromide. Reduction of the azide moiety and subsequent Troc protection produced the disaccharide donor **5** in 81% overall yield for the three steps (Scheme 1). Galactoside **6** and lactoside **7** were prepared according to literature methods.[30,60]

With all necessary building blocks in hand, we performed the assembly of SSEA-3 using the pre-activation-based one-pot protocol.[30,44] With future automa-



Scheme 1.



Scheme 2.

tion in mind, we decided to carry out the one-pot synthesis under the reaction conditions established previously without optimization. Pre-activation of the disaccharide donor **5** at −78 °C with *p*-TolSCl/AgOTf was followed by the addition of the bifunctional building block **6** (Scheme 2, reaction **a**). A sterically hindered base, 2,4,6-tri-*tert*-butylpyrimidine (TTBP)[61] was added with **6** to neutralize triflic acid generated from glycosylation. The reaction temperature was raised to −20 °C to expedite glycosylation, and the acceptor **6** was completely consumed as judged by TLC analysis. The reaction temperature was cooled back down to −78 °C, followed by addition of the lactoside acceptor **7** and *p*-TolSCl/AgOTf. The fully protected SSEA-3 pentasaccharide **4α** was obtained in 37% yield from this three-component one-pot reaction sequence within six hours, which was fully characterized by ^{1}H NMR, ^{13}C NMR, gCOSY, gHMQC and HR-MS. In addition, the anomer **4β** was also separated in 13% yield. The presence or absence of the base TTBP did not significantly affect either the yield or the stereochemical selectivity of this reaction.[62]

Deprotection of the pentasaccharide **4α** was performed by first removing the Troc group with 1 M NaOH in THF followed by acetylation. Staudinger reduction of the azide group and subsequent catalytic hydrogenation over Pearlman's catalyst[30,50,51,63] gave the fully deprotected SSEA-3 **3** in 55% overall yield for all the deprotection steps (Scheme 2, reaction **b**).

Formation of the Challenging α-Gal-(1→4)-Gal Linkage, Stereochemical Dependence on Donor

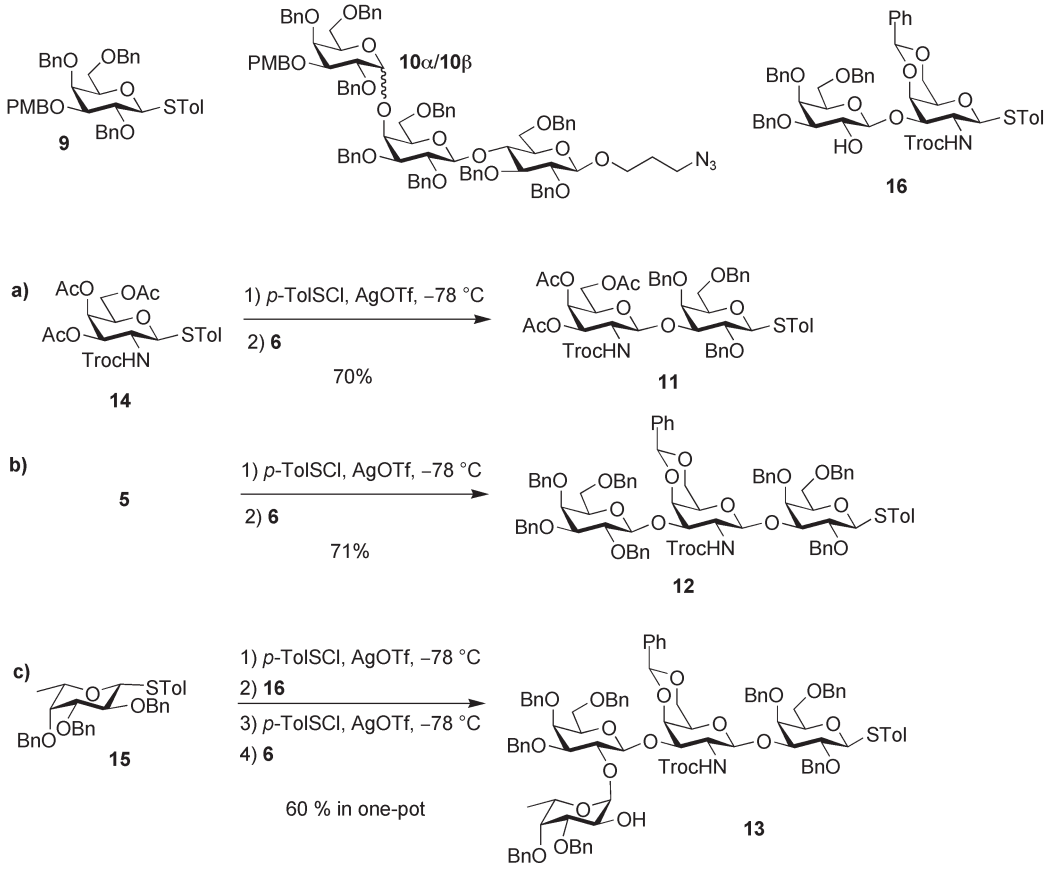
We examined next the stereochemical outcome in formation of the α-Gal-(1→4)-Gal linkage, which is a major challenge in the syntheses of all globo series of gangliosides, with anomeric mixtures of products often obtained.[28,60,64] We have reported that glycosylation of the lactoside acceptor **7** by the galactoside **9**[30] gave the Gb3 trisaccharide **10** in a combined 85% yield with an α:β ratio of 27:1 (Table 1, entry 1),[30] which compared favorably with literature results on Gb3 synthesis.[28,60,64] The large difference in stereochemical selectivity between Gb3 trisaccharide **10** and

Table 1. Evaluation of the formation of the α-Gal-(1→4)-Gal linkage.

Donor (1 equiv.) + AgOTf —[*p*-TolSCl, acceptor **7**, –78 °C; Et_2O:CH_2Cl_2]→ Product

Entry #	Donor	Product[a] (Yield)	α/β
1	**9**	**10**α (82%) + **10**β (3%)	27
2	**11**	**17**α (71%) + **17**β (19%)	3.7
3	**12**	**4**α (60%) + **4**β (23%)	2.6
4	**13**	**18**α (57%) + **18**β (30%)	1.9
5	**19**	**20**α (78%)	α only

[a] Products = **17α/17β**, **18α/18β**, **19**, **20α**



Scheme 3.

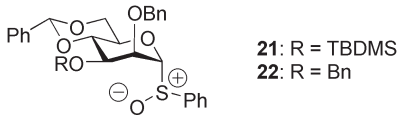
SSEA-3 pentasaccharide **4** prompted us to prepare the disaccharide donor **11**, the trisaccharide donor **12** and the tetrasaccharide donor **13** (Scheme 3). Pre-activation of donor **14**[30] by *p*-TolSCl/AgOTf followed by addition of the galactoside **6** generated the disaccharide **11** in 70% yield and the glycosylation of **6** by **5** produced the trisaccharide **12** (Scheme 3, reactions **a** and **b**). The successful glycosylation of armed galactoside **6** by the disarmed donor **14** is a salient feature of the pre-activation method. One-pot sequential glycosylation of fucoside **15**,[30] disaccharide **16**[30] and galactoside **6** led to the tetrasaccharide donor **13** in 60% overall yield (Scheme 3, reaction **c**).

The glycosylations of lactoside **7** by donors **11**, **12** and **13** were then performed. The reaction of disaccharide donor **11** with **7** gave tetrasaccharides **17α** (71%) and **17β** (19%) (**17α**:**17β** = 3.7:1, Table 1, entry 2). Trisaccharide **12** glycosylated the lactoside **7** with an α:β ratio of 2.6 (Table 1, entry 3), which was similar to that obtained in the one-pot synthesis of SSEA-3 (Scheme 2) indicating that the one-pot procedure did not significantly affect the stereoselectivity. A further reduction in α:β selectivity (1.9:1) was observed in glycosylation of **7** by the tetrasaccharide donor **13** (Table 1, entry 4). These results suggested that as the glycosyl donor became larger, the α selectivity became lower although the overall yields were similar. The stereoselectivity change must be reflective of the rate differential of axial versus equatorial approach of the nucleophile onto the reactive intermediate(s),[51,65] because no anomeric bond isomerizations were observed when the *O*-glycoside products were re-submitted to the glycosylation reaction conditions.

It is possible that the stereochemical divergence in glycosylating the acceptor **7** may be attributed to changes in electronic properties and/or steric encumbrance of the glycosyl donors. The introduction of a single electron-withdrawing group on glycosyl donors can greatly influence the stereoselectivity.[66,67] Inductively, the glycosyl units on 3-O of the reducing end galactose in donors **11**–**13** can be viewed as electron-withdrawing groups.[51,68] Therefore we prepared donor **19** bearing a highly electron-withdrawing *p*-nitrobenzoyl (*p*-NO_2Bz) moiety on its 3-O position through nitrobenzoylation of galactoside **6**. Glycosylation of the lactoside **7** by the donor **19** produced the trisaccharide **20α** in 78% yield with no corresponding β anomer isolated (Table 1, entry 5). This suggests that electronic properties of the donors do not play a

major role in determining stereoselectivity in this case.

Crich and co-workers have reported that the presence of a *tert*-butyldimethylsilyl (TBDMS) group on 3-O of the mannosyl donor **21** led to a significant decrease in β-mannoside formation compared to the corresponding reaction with the benzyl-bearing donor **22**.[69] This was rationalized by a steric buttressing



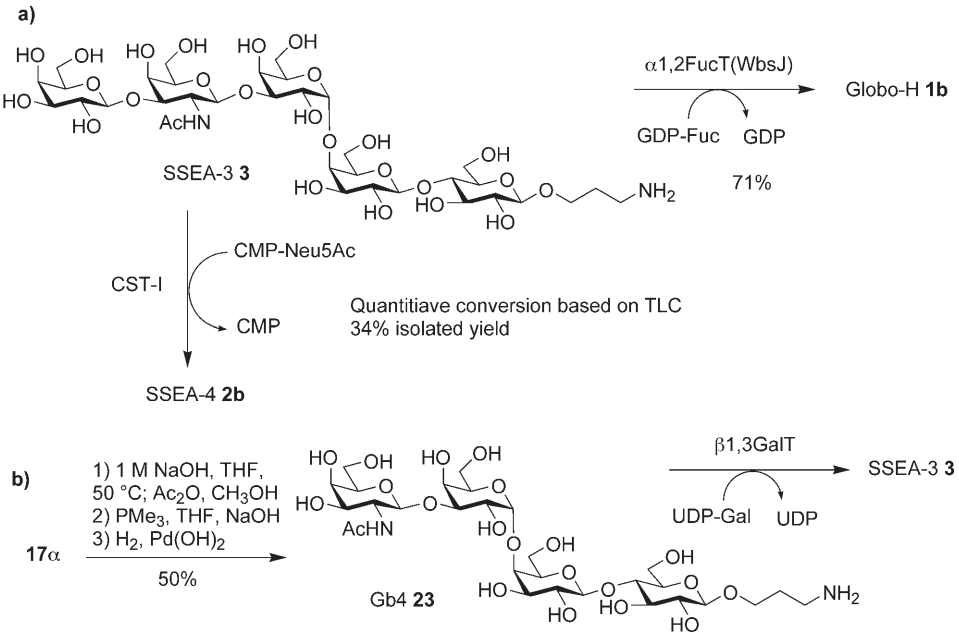
effect as the bulky 3-O-TBDMS moiety pushes the axial 2-O-benzyl group towards the β face of the anomeric center, 1,2-*cis*-(β-mannosyl) glycosylation is hindered. Similar phenomena have been observed in modular synthesis of alternating β-(1→3)-β-(1→4)-mannans,[70] where increased steric size on the 3-O position of the donor in the form of another glycosyl unit reduced the amount of β-mannoside product drastically. It is possible in our studies that the glycosyl units on the 3-O of reducing end galactose of donors **11**, **12** and **13** present a steric buttressing effect, pushing the equatorial 2-O-benzyl towards the anomeric center, thus reducing the amount of 1,2-*cis*-(α-galactosyl) products. As the size of the donor increases from monosaccharide **9** to tetrasaccharide **13**, the effect of the steric buttressing effect becomes greater resulting in lower α selectivity.

Enzymatic Synthesis of Globo-H 1a and MSGG 2a

Fucosylation of the SSEA-3 pentasaccharide **3** was carried out using WbsJ[71] with GDP-fucose. Despite the presence of three galactosyl units in SSEA-3, the 2-OH of the non-reducing terminal galactose was selectively fucosylated leading to Globo-H **1a** in 71% yield (Scheme 4, reaction **a**), the identity of which was confirmed by NMR comparison with the Globo-H **1a** assembled *via* chemical synthesis.[30]

Sialylation of pentasaccharide **3** was performed with the *Campylobacter jejuni* Cst-I sialyltransferase (construct CST-06), which cleanly transformed SSEA-3 into SSEA-4 **2b** with 1.5 equivalents of CMP-Neu5 Ac (Scheme 4, reaction **a**). The conversion was quantitative based on TLC but the isolated yield was 34%. Presumably, some product was lost during the work-up and purification processes. SSEA-3 can also be sialylated by a multifunctional *Pasteurella multocida* sialyltransferase PmST1[72] to form SSEA-4 **2b** in over 90% yield based on TLC analysis. In addition, we found that SSEA-3 is a substrate for a recombinant *Photobacterium damsela* α-2,6-sialyltransferase Pd2,6ST[73] with a yield of 40% by TLC. These results indicate that the large size of the pentasaccharide acceptor **3** does not present an obstacle to these enzymatic glycosylation reactions.

As we have obtained the tetrasaccharide **17α**, we investigated whether its fully deprotected form can also serve as an enzyme substrate. Compound **17α** was deprotected in the same manner as described for the pentasaccharide **3**, producing the tetrasaccharide **23** in 50% yield (Scheme 4, reaction **b**). Tetrasacchar-



Scheme 4.

ide **23** can serve as a substrate to galactosyl transferase LgtD,[41,74] providing an alternative access to SSEA-3.

Conclusions

In conclusion, we report the first chemoenzymatic syntheses of two tumor-associated carbohydrate antigens, Globo-H hexasaccharide **1b** and SSEA-4 hexasaccharide **2b**. The common precursor SSEA-3 pentasaccharide for these two compounds was assembled rapidly using the pre-activation-based one-pot method. Enzymatic glycosylations of SSEA-3 by fucosyltransferase WbsJ and sialyltransferases CST-I or PmST1 led to Globo-H and SSEA-4, respectively. Interestingly, we observed that the stereoselectivities in formation of the challenging α-Gal-(1→4)-Gal linkage decreased when larger oligosaccharide donors were used. This was rationalized by a steric buttressing effect due to the glycosyl unit(s) linked to 3-O of the reducing end galactose in the donor.

Experimental Section

Characterization of Anomeric Stereochemistry

The stereochemistries of the newly formed glycosidic linkages in oligosaccharides are determined by $^3J_{H1,H2}$ through ^{1}H NMR and/or $^1J_{C1,H1}$ through gHMQC 2-D NMR (without ^{1}H decoupling). Smaller coupling constants of $^3J_{H1,H2}$ (around 3 Hz) indicate 1,2-*cis* α linkages and larger coupling constants $^3J_{H1,H2}$ (7.2 Hz or larger) indicate 1,2-*trans* β linkages. This can be further confirmed by $^1J_{C1,H1}$ (~170 Hz) for α linkages and $^1J_{C1,H1}$ (~160 Hz) for β linkages.[75]

General Procedure for Single Step Pre-Activation-Based Glycosylation

A solution of donor (0.060 mmol) and freshly activated molecular sieve MS 4 Å (200 mg) in CH_2Cl_2 (2 mL) was stirred at room temperature for 30 min, and cooled to −78 °C, which was followed by addition of AgOTf (47 mg, 0.18 mmol) dissolved in Et_2O (1 mL) without touching the wall of the flask. After 5 min, orange colored *p*-TolSCl (9.5 μL, 0.060 mmol) was added through a microsyringe. Since the reaction temperature was lower than the freezing point of *p*-TolSCl, *p*-TolSCl was added directly into the reaction mixture to prevent it from freezing on the flask wall. The characteristic yellow color of *p*-TolSCl in the reaction solution dissipated rapidly within a few seconds indicating depletion of *p*-TolSCl. After the donor was completely consumed according to TLC analysis (about 5 min at −78 °C), a solution of acceptor (0.060 mmol) in CH_2Cl_2 (0.2 mL) was slowly added dropwise *via* a syringe. The reaction mixture was warmed to −20 °C under stirring in 2 h. Then the mixture was diluted with CH_2Cl_2 (20 mL) and filtered over Celite. The Celite was further washed with CH_2Cl_2 until no organic compounds were observed in the filtrate by TLC. All CH_2Cl_2 solutions were combined and washed twice with a saturated aqueous solution of $NaHCO_3$ (20 mL) and twice with water (10 mL). The organic layer was collected and dried over Na_2SO_4. After removal of the solvent, the desired disaccharide was purified from the reaction mixture *via* silica gel flash chromatography.

p-Tolyl 2,3,4,6-Tetra-*O*-benzyl-β-D-galactopyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-1-thio-β-D-galactopyranoside (5)

Galactoside **8**[30] (0.4 g, 0.48 mmol) was dissolved in DMF (10 mL) and the solution was cooled to 0 °C. NaH (0.029 g, 60% NaH in mineral oil, 0.72 mmol) was added in portions, followed by addition of benzyl bromide (0.086 mL, 0.72 mmol). The mixture was stirred at room temperature under N_2 for 2 h and then diluted with EtOAc (50 mL). The mixture was washed with saturated $NaHCO_3$, water and then dried over Na_2SO_4, filtered and concentrated. Silica gel column chromatography (2:1 hexanes-EtOAc) afforded *p*-tolyl 2,3,4,6-tetra-*O*-benzyl-β-D-galactopyranosyl-(1→3)-2-azido-4,6-benzylidene-2-deoxy-1-thio-β-D-galactopyranoside **S1** as white solid; yield: 0.44 g (quantitative).

The above prepared **S1** (0.44 g, 0.48 mmol), 1,3-propanedithiol (0.48 mL, 4.8 mmol) and Et_3N (0.67 mL, 4.8 mmol) were dissolved in a mixture of CH_2Cl_2/MeOH (5 mL each). The mixture was heated at reflux overnight under N_2 and then concentrated. The resulting residue was diluted with CH_2Cl_2 (60 mL) and then washed with a saturated aqueous solution of $NaHCO_3$ and water, dried over Na_2SO_4, filtered and concentrated. The resulting residue was purified by quickly passing it through a short silica gel column (20:1, CH_2Cl_2-MeOH) and the obtained solid and solid $NaHCO_3$ (0.078 g, 0.93 mmol) were put into THF (5 mL) and then TrocCl (0.075 mL, 0.56 mmol) was added. The mixture was stirred at room temperature under N_2 for 4 h and filtered. The filtrate was concentrated and then diluted with CH_2Cl_2 (50 mL). The mixture was washed with water and brine, dried over Na_2SO_4, filtered and concentrated. Silica gel column chromatography (2:1 Hexanes-EtOAc) afforded compound **5** as white solid; yield: 0.42 g (81% for two steps).

3-Azidopropyl 2,3,4,6-Tetra-*O*-benzyl-β-D-galactopyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-α-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-glucopyranoside (4α) and 3-Azidopropyl 2,3,4,6-Tetra-*O*-benzyl-β-D-galacto-pyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-trichloro-ethoxycarbonylamino)-2-deoxy-β-D-galacto-pyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-β-D-galacto-pyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-galacto-pyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-glucopyranoside (4β)

After donor **5** (50 mg, 46.7 μmol) and activated molecular sieve MS-4 Å (500 mg) had been stirred for 30 min at room temperature in CH_2Cl_2 (3 mL), the solution was cooled to −78 °C, followed by addition of AgOTf (36 mg, 140 μmol) in

Et_2O (1.5 mL). The mixture was stirred for 5 min at −78 °C and then *p*-TolSCl (7.4 μL, 46.7 μmol) was added into the solution. (See the general procedure for single step pre-activation based glycosylation for precautions) The mixture was vigorously stirred for 10 min, followed by addition of a solution of acceptor **6** (22.1 mg, 39.7 μmol) and TTBP (6.9 mg, 46.7 μmol) in CH_2Cl_2 (1 mL). The reaction mixture was stirred for 2 h from −78 to −20 °C and then the mixture was cooled down to −78 °C, followed by sequential additions of AgOTf (12 mg, 46.7 μmol) in Et_2O (1 mL), acceptor **7** (22.5 mg, 23.3 μmol) and TTBP (6.9 mg, 46.7 μmol) in CH_2Cl_2 (1 mL). The mixture was stirred for 5 min at −78 °C and then *p*-TolSCl (6.3 μL, 39.7 μmol) was added into the solution. The reaction mixture was stirred for 3 h from −78 to 10 °C and then was quenched with Et_3N (40 μL), concentrated under vacuum to dryness. The resulting residue was diluted with CH_2Cl_2 (30 mL), followed by filtration. The organic phase was washed with saturated aqueous $NaHCO_3$, H_2O and then dried over Na_2SO_4, filtered and concentrated. Silica gel column chromatography (2:1 hexanes-EtOAc) afforded **4α** (yield: 20 mg, 37%) and **4β** (yield: 7.2 mg, 13%), respectively, as colorless gel.

Compounds **4α** and **4β** were also synthesized from donor **12** and acceptor **7** in 60% and 23% yield, respectively, as colorless gel following the general procedure of single step glycosylation.

3-Aminopropyl β-D-Galactopyranosyl-(1→3)-2-acetamido-2-deoxy-β-D-galactopyranosyl-(1→3)-α-D-galactopyranosyl-(1→4)-β-D-galactopyranosyl-(1→4)-β-D-glucopyranoside (3)

The mixture of compound **4α** (60 mg, 25.5 μmol), 1 M NaOH (4 mL, 0.4 mmol) and THF (4 mL) was stirred at 50 °C overnight and then concentrated to dryness. The resulting residue was diluted with CH_2Cl_2 (30 mL) and the organic phase was washed by H_2O and then dried over Na_2SO_4, filtered and concentrated to dryness. The resulting residue was dissolved in methanol (2 mL) and acetic anhydride (24 μL, 0.25 mmol) was added dropwise and the mixture was stirred at room temperature under N_2 overnight. The reaction was quenched by adding a few drops of H_2O and then diluted with CH_2Cl_2 (30 mL). The organic phase was washed with a saturated aqueous solution of $NaHCO_3$, H_2O and then dried over Na_2SO_4, filtered and concentrated to dryness. Silica gel column chromatography (2:1 hexanes-EtOAc) afforded the *N*-acetylation product as white solid.

The mixture of the *N*-acetylation product, 1 M PMe_3 in THF (0.5 mL, 0.5 mmol), 0.1 M NaOH (0.5 mL, 0.05 mmol) and THF (4 mL) was stirred at 60 °C under N_2 overnight. The mixture was concentrated and the resulting residue was diluted with CH_2Cl_2 (30 mL). The organic phase was washed with H_2O and then dried over Na_2SO_4, filtered and concentrated to dryness. The resulting residue was purified by quickly passing through a short silica gel column (10:1, CH_2Cl_2-MeOH). The mixture of the obtained solid and $Pd(OH)_2$ in MeOH/H_2O/HOAc (3 mL/1 mL/1 mL) was stirred under H_2 at room temperature overnight and then filtered. The filtrate was concentrated to dryness under vacuum and then was co-evaporated with H_2O (10 mL) three times to remove the HOAc. The aqueous phase was further washed with CH_2Cl_2 (5 mL × 3) and EtOAc (5 mL × 3) and then the aqueous phase was dried under vacuum to afford compound **3** (acetate salt) as a white solid; yield: 13 mg (55% for four steps).

p-Tolyl 3,4,6-tri-*O*-Acetyl-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-1-thio-β-D-galactopyranoside (11)

Compound **11** was synthesized from donor **14** and acceptor **6** in 70% yield following the general procedure of glycosylation.

p-Tolyl 2,3,4,6-Tetra-*O*-benzyl-β-D-galactopyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-1-thio-β-D-galactopyranoside (12)

Compound **12** was synthesized from donor **5** and acceptor **6** in 71% yield following the general procedure of single step glycosylation.

3-Azidopropyl 2,3,4-Tri-*O*-benzyl-α-L-fucopyranosyl-(1→2)-3,4,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-1-thio-β-D-galactopyranoside (13)

Compound **13** was synthesized by a three-component one-pot synthesis procedure. After the donor *p*-tolyl 2,3,4-tri-*O*-benzyl-1-thio-β-L-fucopyranoside **15**[30] (50 mg, 92.47 μmol) and activated molecular sieve MS-4 Å (500 mg) had been stirred for 30 min at room temperature in Et_2O (4 mL), the solution was cooled to −78 °C, followed by addition of AgOTf (72 mg, 277.4 μmol) in Et_2O (1.5 mL). The mixture was stirred for 5 min at −78 °C and then *p*-TolSCl (14.7 μL, 92.47 μmol) was added into the solution. (See the general procedure for single step pre-activation based glycosylation for precautions) The mixture was vigorously stirred for 10 min, followed by addition of a solution of acceptor *p*-tolyl 3,4,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-1-thio-β-D-galactopyranoside **16**[30] (77.1 mg, 78.60 μmol) and TTBP (23 mg, 92.47 μmol) in CH_2Cl_2 (1 mL). The reaction mixture was stirred for 2 h from −78 to −20 °C and then the mixture was cooled down to −78 °C, followed by addition of AgOTf (24 mg, 92.47 μmol) in Et_2O (1 mL). The mixture was stirred for 10 min at −78 °C and then *p*-TolSCl (12.5 μL, 78.60 μmol) was added into the solution. After stirred for 5 min, a solution of acceptor **6** (30.9 mg, 55.48 μmol) and TTBP (23 mg, 92.47 μmol) in CH_2Cl_2 (1 mL) was added slowly along the flask wall into the mixture and the reaction mixture was stirred for 2 h from −78 to −20 °C, then reaction was quenched with Et_3N (40 μL) and concentrated under vacuum to dryness. The resulting residue was diluted with CH_2Cl_2 (30 mL), followed by filtration. The organic phase was washed with saturated aqueous $NaHCO_3$, H_2O and then dried over Na_2SO_4, filtered and concentrated. Silica gel column chromatography (2:1 hexanes-EtOAc) afforded **13** as a colorless gel; yield: 60.6 mg (60%).

3-Azidopropyl 3,4,6-Tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-α-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-β-D-glucopyranoside (17α) and 3-Azidopropyl 3,4,6-Tri-*O*-acetyl-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-β-D-glucopyranoside (17β)

Compound **17α** and **17β** were synthesized from donor **11** and acceptor **7** in 71% and 19% yield, respectively, as colorless gel following the general procedure of single-step glycosylation.

3-Azidopropyl 2,3,4-Tri-*O*-benzyl-α-L-fucopyranosyl-(1→2)-3,4,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-trichloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-α-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-glucopyranoside (18α) and 3-Azidopropyl 2,3,4-Tri-*O*-benzyl-α-L-fucopyranosyl-(1→2)-3,4,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→3)-4,6-benzylidene-2-(2,2,2-tri-chloroethoxycarbonylamino)-2-deoxy-β-D-galactopyranosyl-(1→3)-2,4,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-glucopyranoside (18β)

Compound **18α** and **18β** were synthesized from donor **13** and acceptor **7** in 57% and 30% yield, respectively, as colorless gel following the general procedure of single step glycosylation. Their identities were confirmed by NMR comparison with literature data.[30]

***p*-Tolyl 2,4,6-Tri-*O*-benzyl-3-*O*-*p*-nitrobenzoyl-1-thio-β-D-galactopyranoside (19)**

Compound **6** (43.5 mg, 0.078 mmol) and *N,N*-dimethylaminopyridine (9.5 mg, 0.078 mmol) were dissolved in CH_2Cl_2 (5 mL), followed by the addition of *p*-nitrobenzoyl chloride (10.9 μL, 0.09 mmol). The mixture was stirred overnight at room temperature and then diluted with CH_2Cl_2 (20 mL), washed with saturated aqueous $NaHCO_3$ and H_2O and then dried over Na_2SO_4, filtered and concentrated. Silica gel column chromatography (4:1 hexanes-EtOAc) afforded **19** as a gel-like solid; yield: 50.6 mg (92%).

3-Azidopropyl 2,4,6-Tri-*O*-benzyl-3-*O*-*p*-nitrobenzoyl-α-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-galactopyranosyl-(1→4)-2,3,6-tri-*O*-benzyl-β-D-glucopyranoside (20α)

After donor **19** (30 mg, 42 μmol), acceptor **7** (37 mg, 38 μmol) and activated molecular sieve MS-4 Å (200 mg) were stirred for 30 min at room temperature in a mixture solvent of Et_2O (1 mL) and CH_2Cl_2 (2 mL), the mixture was cooled to −78 °C, followed by addition of AgOTf (33 mg, 0.127 mmol) in Et_2O (1 mL). The mixture was vigorously stirred for 10 min and then *p*-TolSCl (6.74 μL, 42 μmol) was added and the reaction mixture was stirred for 2 h from −78 to −20 °C. The reaction was quenched by Et_3N and then concentrated under vacuum to dryness. The resulting residue was diluted with CH_2Cl_2 (10 mL), followed by filtration. The organic phase was washed with saturated aqueous $NaHCO_3$ and H_2O and then dried over Na_2SO_4, filtered and concentrated. Silica gel column chromatography (3:1 hexanes-EtOAc) afforded **20α** as gel-like solid; yield: 46.2 mg (78%).

3-Aminopropyl α-L-Fucopyranosyl-(1→2)-β-D-galactopyranosyl-(1→3)-2-acetamido-2-deoxy-β-D-galac-topyranosyl-(1→3)-α-D-galactopyranosyl-(1→4)-β-D-galactopyranosyl-(1→4)-β-D-glucopyranoside (1b)

To a mixture of 10 mM SSEA-3 pentasaccharide **3**, 15 mM GDP-fucose in 20 mM Tris-HCl buffer (pH 7.5) was added α-1,2-fucosyltransferase WbsJ (7 mU). The mixture was incubated at room temperature for 2 days. The Globo-H hexasaccharide **1b** was purified on a BioGel P-2 gel filteration column (BioRad) followed by Dowex 1×8–400 ion exchange resin (chloride form) with water as the mobile phase. The fractions containing the desired product **1b** (71%) were collected and lyophilized. NMR data of **1b** were collected, which were identical to those obtained from Globo-H **1b** synthesized chemically.[30]

3-Aminopropyl 5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-non-2-ulopyranosylonic acid-(2→3)-β-D-galactopyranosyl-(1→3)-2-acetamido-2-deoxy-β-D-galactopyranosyl-(1→3)-α-D-galactopyranosyl-(1→4)-β-D-galactopyranosyl-(1→4)-β-D-glucopyranoside (2b)

The *Campylobacter jejuni* CST-I α-2,3-sialyltransferase (construct CST-06) was expressed as a fusion protein with the *E. coli* maltose-binding protein and purified.[39] To a mixture of 10 mM $MgCl_2$, 5 mM SSEA-3 pentasaccharide **3**, 7.5 mM CMP-Neu5Ac in Hepes buffer (50 mM, pH 7.5) was added CST-I α-2,3-sialyltransferase (0.32 U/mL). The reaction was incubated at 37 °C for 2 h and TLC (BPH:AMW = 1:1, BPH 1:2:1 *n*-butanol:propanol:0.1 M HCl; AMW 1:1:1 CH_3CN:methanol:water) showed complete consumption of the pentasaccharide **3**. Multiple reactions were set to convert all of the pentasaccharide **3** (9 mg, 9.1 μmol). The reaction volume was reduced using a centrifugal concentrator and the material was loaded (in 3 runs) on a 10 mm × 30 cm Superdex Peptide column (GE Healthcare). The column was eluted with an ammonium acetate buffer (pH 7, 20 mM) at 0.5 mL min^{-1}. The product was eluted between 7 and 8 mL. The fractions were pooled and lyophilized three times to remove the ammonium acetate. We recovered 4 mg (3.1 μmol) of purified **2b** with a yield of 34%. As the conditions for enzymatic synthesis were very mild and previously we did not observe any product decomposition using this enzyme,[39] the low isolated yield was probably due to multiple small-scale reactions performed resulting in loss of product during work-up and purification.

Enzymatic Sialylation of Pentasaccharide SSEA-3 using PmST1 and Pd2,6ST

The enzymatic assays were carried out in a total volume of 10 μL in a Tris-HCl buffer (100 mM, pH 8.5) containing CMP-Neu5Ac (20 mM), SSEA-3 **3** (10 mM), $MgCl_2$ (20 mM) and the corresponding sialyltransferases (PmST1 or Pd2,6ST). Reactions were allowed to proceed for 60 min at 37 °C. The reactions were then monitored by TLC (BPH: AMW = 1:1) and the products formed were confirmed by MS. ESI-MS (for sialylation product catalyzed by PmST1): m/z = 1262.29 calcd. for $[M+Na]^+$ $C_{46}H_{78}N_3Na_2O_{34}$: 1262.43; ESI-MS (for sialylation product catalyzed by Pd2,6ST): m/z = 1262.28, calcd. for $[M+Na]^+$ $C_{46}H_{78}N_3Na_2O_{34}$: 1262.43.

3-Aminopropyl 2-Acetamido-2-deoxy-β-D-galactopyranosyl-(1→3)-α-D-galactopyranosyl-(1→4)-β-D-galactopyranosyl-(1→4)-β-D-glucopyranoside (23)

The mixture of compound **17α** (0.050 g, 0.027 mmol), 1 M NaOMe (4.0 mL, 4.0 mmol) and THF (4 mL) was stirred at 50 °C overnight and then concentrated to dryness. The resulting residue was diluted with CH_2Cl_2 (50 mL) and the organic phase was washed by H_2O and then dried over Na_2SO_4, filtered and concentrated to dryness. The resulting residue was dissolved in methanol (3 mL). Acetic anhydride (1.0 mL) was added dropwise and the mixture was stirred at room temperature under N_2 for 6 h. The reaction was quenched by adding EtOH and then diluted with EtOAc (20 mL). The organic phase was washed with saturated aqueous solution of $NaHCO_3$, H_2O and then dried over Na_2SO_4, filtered and concentrated to dryness. Silica gel column chromatography (2:1 hexanes-EtOAc) afforded the *N*-acetylation product as white solid.

The mixture of the *N*-acetylation product, 1 M PMe_3 in THF (0.5 mL, 0.5 mmol) and THF (3 mL) was stirred at 50 °C under N_2 overnight. The mixture was concentrated and the resulting residue was diluted with CH_2Cl_2 (50 mL). The organic phase was washed with H_2O and then dried over Na_2SO_4, filtered and concentrated to dryness. The resulting residue was purified by quickly passing through a short silica gel column (10:1, CH_2Cl_2-MeOH). The mixture of the obtained solid and $Pd(OH)_2$ in MeOH/H_2O/HOAc (5 mL, 3:1:1) was stirred under H_2 at room temperature overnight and then filtered. The filtrate was concentrated to dryness under vacuum and then was co-evaporated with H_2O (10 mL) three times to remove the HOAc. The aqueous phase was further washed with CH_2Cl_2 (5 mL × 3) and EtOAc (5 mL × 3) and then the aqueous phase was dried under vacuum to afford compound **23** as white solid; yield: 10.4 mg (50% for three steps).

Enzymatic Galactosylation of Tetrasaccharide 23

To a mixture of 10 mM tetrasaccharide **23**, 20 mM UDP-galactose, 1 mM $MnCl_2$, 1 mM dithiothreitol and 1% BSA in 20 mM Tris-HCl buffer (pH 7.5) was added β-1,3-galactosyltransferase (LgtD) (2 mU). The mixture was incubated at room temperature for 2 days. The SSEA-3 pentasaccharide **3** (70%) was purified on a BioGel P-2 gel filteration column (BioRad) followed by Dowex 1×8–400 ion exchange resin (chloride form) with water as the mobile phase.

Supporting Information

General experimental procedures as well as analytical and spectroscopic characterization data for all compounds are given in the Supporting information.

Acknowledgements

We are grateful for financial supports from the National Institutes of Health (R01-GM-72667 to XH, R01-GM-76360 to XC). We would like to thank Dr. Warren Wakarchuk (National Research Council Canada) for helpful suggestions, Dr. Jianjun Li (National Research Council Canada) for mass spectrometry analysis of the sialylated product and Marie-France Karwaski (National Research Council Canada) for technical help.

References

[1] H. Ishida, M. Kiso, *Trends Glycosci. Glycotechnol.* **2001**, *13*, 57–64.
[2] S. Hakomori, Y. Zhang, *Chem. Biol.* **1997**, *4*, 97–104.
[3] A. Wedeking, G. van Echten-Deckert, *Curr. Org. Chem.* **2007**, *11*, 579–598.
[4] M. Izumi, H. Uzawa, *Trends Glycosci. Glycotechnol.* **2005**, *17*, 107–119.
[5] S. Birkle, G. Zeng, L. Gao, R. K. Yu, J. Aubry, *Biochimie* **2003**, *85*, 455–463.
[6] S. Zhang, H. S. Zhang, V. E. Reuter, S. F. Slovin, H. I. Scher, P. O. Livingston, *Clin. Cancer Res.* **1998**, *4*, 295–302.
[7] E. G. Bremer, S. B. Levery, S. Sonnino, R. Ghidoni, S. Canevari, R. Kannagi, S. Hakomori, *J. Biol. Chem.* **1984**, *259*, 14773–14777.
[8] S. J. Danishefsky, J. R. Allen, *Angew. Chem.* **2000**, *112*, 882–912; *Angew. Chem. Int. Ed.* **2000**, *39*, 836–863.
[9] T. Gilewske, G. Ragupathi, S. Bhuta, L. J. Williams, C. Musselli, X.-F. Zhang, K. P. Bencsath, K. S. Panageas, J. Chin, C. A. Hudis, L. Norton, A. N. Houghton, P. O. Livingston, S. J. Danishefsky, *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 3270–3275.
[10] Z. G. Wang, L. J. Williams, X. F. Zhang, A. Zatorski, V. Kudrya-shov, G. Ragupathi, M. Spassova, W. Bornmann, S. F. Slovin, H. I. Scher, P. O. Livingston, K. O. Lloyd, S. J. Danishefsky, *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 2719.
[11] S. F. Slovin, G. Ragupathi, S. Adluri, G. Ungers, K. Terry, S. Kim, M. Spassova, W. G. Bornmann, M. Fazzari, L. Dantis, K. Olkiewicz, K. O. Lloyd, P. O. Livingston, S. J. Danishefsky, H. I. Scher, *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 5710.
[12] A. E. Stapleton, M. R. Stroud, S. I. Hakomori, W. E. Stamm, *Infect. Immun.* **1998**, *66*, 3856–3861.
[13] M. R. Stroud, A. E. Stapleton, S. B. Levery, *Biochemistry* **1998**, *37*, 17420–17428.
[14] A. Stapleton, E. Nudelman, H. Clausen, S. Hakomori, W. E. Stamm, *J. Clin. Invest.* **1992**, *90*, 965–972.
[15] L. L. W. Cooling, T. A. W. Koerner, S. J. Naides, *J. Infect. Dis.* **1995**, *172*, 1198–1205.

[16] Y. U. Katagiri, K. Ohmi, C. Katagiri, T. Sekino, H. Nakajima, T. Ebata, N. Kiyokawa, J. Fujimoto, *Glycoconjugate J.* **2001**, *18*, 347–353.
[17] J. Wenk, P. W. Andrews, J. Casper, J. Hata, M. F. Pera, A. von Keitz, I. Damjanov, B. A. Fenderson, *Int. J. Cancer* **1994**, *58*, 108–115.
[18] J. G. Krupnick, I. Damjanov, A. Damjanov, Z. M. Zhu, B. A. Fenderson, *Int. J. Cancer* **1994**, *59*, 692–698.
[19] R. Kannagi, S. B. Levery, F. Ishigami, S. Hakomori, L. H. Shevinsky, B. B. Knowles, D. Solter, *J. Biol. Chem.* **1983**, *258*, 8934–8942.
[20] S. Saito, H. Aoki, A. Ito, S. Ueno, T. Wada, K. Mitsuzuka, M. Satoh, Y. Arai, T. Miyagi, *J. Biol. Chem.* **2003**, *278*, 26474–26479.
[21] W. F. Steelant, Y. Kawakami, A. Ito, K. Handa, E. A. Bruyneel, M. Mareel, S. Hakomori, *FEBS Lett.* **2002**, *531*, 93–98.
[22] S. Saito, S. Orikasa, M. Satoh, C. Ohyama, A. Ito, T. Takahashi, *Jpn. J. Cancer Res.* **1997**, *88*, 652–659.
[23] S. Saito, S. Orikasa, C. Ohyama, M. Satoh, Y. Fukushi, *Int. J. Cancer* **1991**, *49*, 329–334.
[24] M. Kiso, H. Ishida, A. Hiromne, in: *Carbohydrate-Based Drug Discovery*, Vol. 1, (Ed.: C.-H. Wong), Wiley-VCH, Weinheim, **2003**, pp 37–54.
[25] J. R. Allen, J. G. Allen, X.-F. Zhang, L. J. Williams, A. Zatorski, G. Ragupathi, P. O. Livingston, S. J. Danishefsky, *Chem. Eur. J.* **2000**, *6*, 1366–1375.
[26] J. M. Lassaletta, R. R. Schmidt, *Liebigs Ann. Chem.* **1996**, 1417–1423.
[27] T. Zhu, G. J. Boons, *Angew. Chem.* **1999**, *111*, 3704–3707; *Angew. Chem. Int. Ed.* **1999**, *38*, 3495–3497.
[28] D. D. Werz, B. Castagner, P. H. Seeberger, *J. Am. Chem. Soc.* **2007**, *129*, 2770–2771.
[29] C.-Y. Huang, D. A. Thayer, A. Y. Chang, M. Best, L. Hoffmann, S. Head, C.-H. Wong, *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 15–20.
[30] Z. Wang, L. Zhou, K. El-boubbou, X.-S. Ye, X. Huang, *J. Org. Chem.* **2007**, *72*, 6409–6420.
[31] D. Crich, B. Wu, *Tetrahedron* **2008**, *64*, 2042–2047 and references cited therein.
[32] G. J. Boons, A. Demchenko, *Chem. Rev.* **2000**, *100*, 4539–4565.
[33] J. Seifert, M. Lergenmüller, Y. Ito, *Angew. Chem.* **2000**, *112*, 541–544; *Angew. Chem. Int. Ed.* **2000**, *39*, 531–534.
[34] H. Ishida, R. Miyawaki, M. Kiso, A. Hasegawa, *J. Carbohydr. Chem.* **1996**, *15*, 163–182.
[35] J. M. Lassaletta, K. Carlsson, P. J. Garegg, R. R. Schmidt, *J. Org. Chem.* **1996**, *61*, 6873–6880.
[36] K. M. Koeller, C.-H. Wong, *Chem. Rev.* **2000**, *100*, 4465–4493, and references cited therein.
[37] N. Wymer, E. J. Toone, *Curr. Opin. Chem. Biol.* **2000**, *4*, 110–119.
[38] R. I. Duclos, *Carbohydr. Res.* **2000**, *328*, 489–507.
[39] O. Blixt, D. Vasiliu, K. Allin, N. Jacobsen, D. Warnock, N. Razi, J. C. Paulson, S. Bernatchez, M. Gilbert, W. Wakarchuk, *Carbohydr. Res.* **2005**, *340*, 1963–1972.
[40] V. P. Kamath, R. E. Yeske, J. M. Gregson, R. M. Ratcliffe, Y. R. Fang, M. M. Palcic, *Carbohydr. Res.* **2004**, *339*, 1141–1146, and references cited therein.
[41] M. Randriantsoa, S. Drouillard, C. Breton, E. Samain, *FEBS Lett.* **2007**, *581*, 2652–2656.
[42] R. Kannagi, N. A. Cochran, F. Ishigami, S. Hakomori, P. W. Andrews, B. B. Knowles, D. Solter, *EMBO J.* **1983**, *2*, 2355–2361.
[43] D. R. Mootoo, P. Konradsson, U. Udodong, B. Fraser-Reid, *J. Am. Chem. Soc.* **1988**, *110*, 5583–5584.
[44] X. Huang, L. Huang, H. Wang, X.-S. Ye, *Angew. Chem.* **2004**, *116*, 5333–5336; *Angew. Chem. Int. Ed.* **2004**, *43*, 5221–5224.
[45] J. D. C. Codée, L. J. van den Bos, R. E. J. N. Litjens, H. S. Overkleeft, C. A. A. van Boeckel, J. H. van Boom, G. A. van der Marel, *Tetrahedron* **2004**, *60*, 1057–1064.
[46] S. Yamago, T. Yamada, T. Maruyama, J.-I. Yoshida, *Angew. Chem.* **2004**, *116*, 2197–2200; *Angew. Chem. Int. Ed.* **2004**, *43*, 2145–2148.
[47] H. M. Nguyen, J. L. Poole, D. Y. Gin, *Angew. Chem.* **2001**, *113*, 428–431; *Angew. Chem. Int. Ed.* **2001**, *40*, 414–417.
[48] D. Crich, S. Sun, *Tetrahedron* **1998**, *54*, 8321–8348.
[49] L. Huang, X. Huang, *Chem. Eur. J.* **2007**, *13*, 529–540.
[50] A. Miermont, Y. Zeng, Y. Jing, X.-S. Ye, X. Huang, *J. Org. Chem.* **2007**, *72*, 8958–8961.
[51] N. Teumelsan, X. Huang, *J. Org. Chem.* **2007**, *72*, 8976–8979.
[52] L. Huang, Z. Wang, X. Li, X.-S. Ye, X. Huang, *Carbohydr. Res.* **2006**, *341*, 1669–1679.
[53] L. Huang, Z. Wang, X. Huang, *Chem. Commun.* **2004**, *17*, 1960–1961.
[54] V. Martichonok, G. M. Whitesides, *J. Org. Chem.* **1996**, *61*, 1702–1706.
[55] D. Crich, M. Smith, *J. Am. Chem. Soc.* **2001**, *123*, 9015–9020.
[56] D. Crich, M. Smith, *Org. Lett.* **2000**, *2*, 4067–4069.
[57] C. Wang, H. Wang, X. Huang, L.-H. Zhang, X.-S. Ye, *Synlett* **2006**, 2846–2850.
[58] J. D. C. Codée, R. E. J. N. Litjens, R. den Heeten, H. S. Overkleeft, J. H. van Boom, G. A. van der Marel, *Org. Lett.* **2003**, *5*, 1519–1522.
[59] J. D. C. Codée, L. J. van den Bos, R. E. J. N. Litjens, H. S. Overkleeft, J. H. van Boom, G. A. van der Marel, *Org. Lett.* **2003**, *5*, 1947–1950.
[60] P. Pornsuriyasak, A. V. Demchenko, *Carbohydr. Res.* **2006**, *341*, 1458–1466, and references cited therein.
[61] D. Crich, M. Smith, Q. Yao, J. Picione, *Synthesis* **2001**, 323–326.
[62] Y. Gen, L.-H. Zhang, X.-S. Ye, *Chem. Commun.* **2008**, 597–599.
[63] J. Li, J. Wang, P. G. Czyryca, H. Chang, T. W. Orsak, R. Evanson, C.-W. T. Chang, *Org. Lett.* **2004**, *6*, 1381–1384.
[64] T. Y. Park, I. J. Kim, S. Hu, M. T. Bilodeau, J. T. Randolph, O. Kwon, S. J. Danishefsky, *J. Am. Chem. Soc.* **1996**, *118*, 11488–11500.
[65] B. Garcia, D. Y. Gin, *J. Am. Chem. Soc.* **2000**, *122*, 4269–4279.
[66] A. A.-H. Abdel-Rahman, S. J. Jonke, E. S. H. El Ashry, R. R. Schmidt, *Angew. Chem.* **2004**, *116*, 4489; *Angew. Chem. Int. Ed.* **2004**, *43*, 4389.
[67] D. Crich, J. Picione, *Org. Lett.* **2003**, *5*, 781–784.
[68] Z. Zhang, I. R. Ollman, X.-S. Ye, R. Wischnat, T. Baasov, C.-H. Wong, *J. Am. Chem. Soc.* **1999**, *121*, 734–753.

[69] D. Crich, V. Dudkin, *Tetrahedron Lett.* **2000**, *41*, 5643–5646.
[70] D. Crich, W. Li, H. Li, *J. Am. Chem. Soc.* **2004**, *126*, 15081–15086.
[71] W. Yi, J. Shao, L. Zhu, M. Li, M. Singh, Y. Lu, S. Lin, H. Li, K. Ryu, J. Shen, H. Guo, Q. Yao, C. A. Bush, P. G. Wang, *J. Am. Chem. Soc.* **2005**, *127*, 2040–2041.
[72] H. Yu, H. Chokhawala, H. Karpel, H. Yu, B. Wu, J. Zhang, Y. Zhang, Q. Jia, X. Chen, *J. Am. Chem. Soc.* **2005**, *127*, 17618–17619.
[73] H. Yu, S. Huang, H. Chokhawala, M. Sun, H. Zheng, X. Chen, *Angew. Chem. Int. Ed.* **2005**, *44*, 3938–3944.
[74] D. M. Su, H. Eguchi, W. Yi, L. Li, P. G. Wang, C. Xia, *Org. Lett.* **2008**, *10*, in press.
[75] K. Bock, C. Pedersen, *J. Chem. Soc. Perkin Trans. 2* **1974**, 293–297.

